

Product datasheet

AAV6 VP1, recombinant protein

Short overview

Cat. No.	640836
Quantity	10 µg
Concentration	100 μg/ml (1.19 μM)

Product description

Formulation	Liquid, 6 M urea in PBS
Source	Escherichia coli
Molecular Weight	83.6 kDa (calculated Mw from aa sequence)
Purity	> 95% (determined by SDS PAGE)
Product description	N-terminal His-tagged (MGSSHHHHHHSSGLVPRGSH) recombinant AAV6 capsid protein VP1
Purification	Ni-NTA chromatography
Storage	-80°C
Intended use	Research use only
Application	Dot blot, SDS PAGE, WB

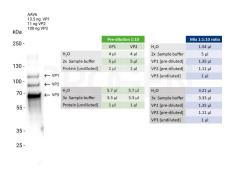
Applications

Dot Blot	100 ng, depending on primary antibody and detection method
SDS PAGE	1 µg
Western Blot (WB)	5-20 ng, depending on primary antibody and detection method

Background

The AAV capsid consists of three capsid proteins, i.e. VP1, VP2 and VP3, which differ in their N-terminus and encapsulate the genomic ssDNA. In native virus particles, the three proteins form subunits with a ratio of 1:1:10 (VP1:VP2:VP3), in a total number of 60 subunits per capsid. The recombinant AAV6 VP1 protein in combination with recombinant AAV6 VP2 (Cat. No. 640837) and recombinant AAV6 VP3 (Cat. No. 640838) can be used to create a mixture with the precise molar ratio of 1:1:10 to compare the protein composition of the viral capsid in your sample by protein detection methods, e.g. western blot. All three recombinant AAV6 capsid proteins are available as set (Cat. No. 72006) or as individual proteins (Cat. No. 640836, 640837, 640838). Note: please find an example how to prepare western blot samples in the pipetting scheme below. Aliquots of the remaining samples can be stored at -80°C for reuse.

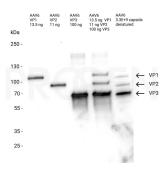
Product images



Pipetting scheme for western blot analysis using a mix of the AAV6 capsid proteins (Cat. No. 640836, 640837, 640838). To create a VP mixture with the molar ratio 1:1:10 (VP1:VP2:VP3), please pre-dilute VP1 and VP2 1:10 to yield a final concentration of 10 µg/ml (green table). Pipette the pre-diluted VP1 and VP2 proteins and mix them with the undiluted VP3 protein in your sample buffer and water (blue table). The example with 2x and 3x sample buffer and the required volumes are indicated in the pipetting scheme. Thus, in one lane, 10 µl of the VP mix can be loaded onto the SDS PAGE and analyzed by Western blot using the B1 antibody (Cat. No. 690058, Cat. No. 61058-488, Cat. No. 61058-647).Undiluited = 100 µg/ml, pre-diluted = 10 µg/ml

kDa	AAV6 VP1 13.5 ng	AAV6 VP2 11 ng	AAV6 VP3 100 ni	g				
250 ·						VP1	VP2	VP3
130 -					H ₂ O	48.65 µl	48.89 µl	4 µl
100	-			← VP1	2x Sample buffer	50 µl	50 µl	5 µl
100 -	-	-		← VP2	Protein [100 µg/ml]	1.35 µl	1.11 µl	1 μΙ
70			-	← VP3				
55			100		H ₂ O	65.35 µl	65.59 µl	5.7 µl
					3x Sample buffer	33.3 µl	33.3 µl	3.3 µl
35					Protein [100 µg/ml]	1.35 µl	1.11 µl	1 µl
25								

Pipetting scheme for western blot analysis using the AAV6 capsid proteins (Cat. No. 640836, 640837, 640838) in separate lanes. To analyze the molar ratio of 1:1:10, it is recommended to load VP1, VP2 and VP3 as described in the pipetting scheme above. Therefore, the indicated volumes of the proteins (concentration 100 μ g/ml) should be diluted with the appropriate amount of sample buffer and distilled water. 10 μ l of each solution can be separately loaded onto the SDS PAGE and analyzed by Western blot using the B1 antibody (Cat. No. 690058, Cat. No. 61058-488, Cat. No. 61058-647).



Western blot analysis of recombinant AAV6 capsid proteins (Cat. No. 640836, 640837, 640838) and denatured AAV6 capsids with B1 antibody (Cat. No. 690058). Western blot analysis was performed on the precise molar ratio of 1:1:10 (VP1:VP2:VP3) either in separate lanes or combined in one lane and on 3.3E+09 denatured AAV6 capsids. The PVDF membrane was blocked with 5% milk in PBST for 1 h at RT. The primary antibody anti-AAV VP1/VP2/VP3, B1 (Cat. No. 690058) was diluted in blocking buffer (antibody concentration 500 ng/ml) and incubated for 1 h at RT. The secondary antibody anti-mouse IgG HRP was also diluted in blocking buffer (antibody concentration 200 ng/ml) and incubated for 1 h at RT. The bands were visualized by chemiluminescent detection using Pierce ECL Western Blotting Substrate.