

## **Product datasheet**

# anti-AAV5 (intact particle) mouse monoclonal, ADK5b, lyophilized, purified

#### Short overview

 Cat. No.
 610149

 Quantity
 50 μg

Concentration 50 µg/ml after reconstitution with 1 ml dist. water

### **Product description**

Host Mouse
Antibody Type Monoclonal
Isotype IgG2b kappa
Clone ADK5b
Immunogen AAV5 capsids

Formulation Lyophilized; reconstitute in 1 ml dist. water (final solution contains 0.09% sodium azide, 0.5% BSA

in PBS buffer, pH 7.4)

**Binding affinity** KD value (AAV5) = 6.0E-11 M **Synomym** Adeno-associated Virus 5, AAV-5

Conjugate Unconjugated

**Purification** Affinity chromatography

**Storage before** 2-8°C until indicated expiry date

reconstitution

Storage after Up to 3 months at 2-8°C; long term storage in aliquots at -20°C; avoid freeze/thaw cycles

reconstitution

Intended use Research use only

**Application** Dot blot, ELISA, ICC/IF, IP, Neutralization assay

Reactivity AAV5

No reactivity AAV1, AAV2, AAV3, AAV4, AAV6, AAV8, AAV9, AAVDJ, AAVrh10, AAVrh74

## **Applications**

**Dot Blot** 1:500 (0.1 μg/ml; non-denaturing conditions)

**ELISA** Assay dependent

Immunocytochemistry (ICC) 1:20
Immunoprecipitation (IP) 1:5

Neutralization Assay EC50 ~9 ng/ml (AAV5) - assay dependent

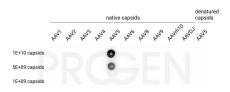
## Background

For characterization of different stages of infection and very useful for the analysis of the AAV5 assembly process. ADK5b specifically reacts with intact adeno-associated virus 5 particles, empty and full capsids. Recognizes a conformational epitope of assembled capsids, not present in PROGEN Biotechnik GmbH | Maaßstraße 30 | D-69123 Heidelberg

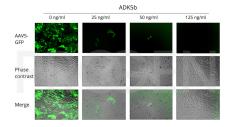
denatured capsid proteins and native but unassembled capsid proteins. The antibody cannot be used for immunoblotting. The antibody is usefull for neutralizing experiments.

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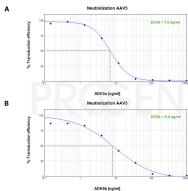
#### **Product images**



Dot blot analysis of native AAV1-AAV9, AAVrh10, AAVDJ capsids (1E+09-1E+10 capsids) and denatured AAV5 capsids (1E+09-1E+10 capsids, denatured at 95°C for 10 min in sample buffer). The nitrocellulose membrane was blocked with 5% dry milk in PBST (PBS + 0.1% Tween 20) for 1 h at RT. The primary antibody anti-AAV5 (intact particle) mouse monoclonal, ADK5b (Cat. No. 610149) was diluted in blocking buffer (antibody concentration 100 ng/ml) and incubated for 1 h at RT. The secondary antibody goat anti-mouse IgG HRP was also diluted in blocking buffer (antibody concentration 200 ng/ml) and incubated for 1 h at RT. The bands were visualized by chemiluminescent detection using PierceTM ECL Plus Western Blotting Substrate.



Neutralization of AAV5-GFP vectors with the ADK5b antibody (Cat. No. 610149). AAV infection was shown in HeLa cells and photos (GFP, CPE, merge) were taken ~48 h post infection. Neutralization was enhanced with increasing ADK5b concentration.



Neutralization of AAV5 with mouse monoclonal AAV5 antibody clone ADK5a (A) and mouse monoclonal AAV5 antibody clone ADK5b (B) by using AAV5-NanoLuc® viral particles from Promega. (A) anti-AAV5 (intact particle) mouse monoclonal, ADK5a (Cat. No. 610148) or (B) anti-AAV5 (intact particle) mouse monoclonal, ADK5b (Cat. No. 610149) were preincubated with AAV5-NanoLuc® viral particles for 30 min at RT at 300 rpm (antibody concentrations 0.2-3,000 ng/ml). HEK293 cells (100 µl) were plated at 200,000 cells/ml in DMEM + 1% FCS. Virus-antibody-mix (20 µl) was added to the cells and incubated for 16-24 h at 37°C. Extracellular NanoLuc Inhibitor and Nano-Glo® Live Cell Assay System (Promega) was added to the wells and incubated for 5 min at RT at 300 rpm. Luminescence was measured using an ID5-Reader and plotted with Softmax Pro 7.1 PROGEN Biotechnik GmbH | Maaßstraße 30 | D-69123 Heidelberg

software to determine the EC50 values.

# References

Publication	Species	Application
Silveria, M. A., Large, E. E., Zane, G. M., White, T. A. & Chapman, M. S. The structure of an aav5-aavr complex at 2.5 Ã resolution: Implications for cellular entry and immune neutralization of aav gene therapy vectors. Viruses 12, (2020).	AAV5	neutralization
Emmanuel, S. N., Mietzsch, M., Tseng, Y. S., Smith, J. K. & Agbandje-Mckenna, M. Parvovirus Capsid-Antibody Complex Structures Reveal Conservation of Antigenic Epitopes across the Family. Viral Immunol. 34, 3–17 (2021).	AAV5	binding region
Emmanuel, S. N. et al. Structurally Mapping Antigenic Epitopes of Adeno-associated Virus 9: Development of Antibody Escape Variants. J. Virol. 96, (2022).	AAV5	dot blot
Jose, A. et al. High-Resolution Structural Characterization of a New Adeno-associated Virus Serotype 5 Antibody Epitope toward Engineering Antibody-Resistant Recombinant Gene Delivery Vectors. J. Virol. 93, 1394–1412 (2019).	AAV5	cryoEM, dot blot
Kuck, D., Kern, A. & Kleinschmidt, J. A. Development of AAV serotype-specific ELISAs using novel monoclonal antibodies.  J. Virol. Methods 140, 17–24 (2007).	AAV5	dot blot,IP