Product datasheet

anti-AAV2 (intact particle) mouse monoclonal, A20, lyophilized, purified, sample

Short overview

Cat. No.	61055S
Quantity	10 µg
Concentration	50 µg/ml after reconstitution with 200 µl dist. water

Product description

Host	Mouse
Antibody Type	Monoclonal
Isotype	IgG3
Clone	A20
Immunogen	AAV2 capsids
Formulation	Lyophilized; reconstitute in 200 µl dist. water (final solution contains 0.09% sodium azide, 0.5% BSA in PBS buffer, pH 7.4)
Binding affinity	KD value (AAV2) = 2.6E-11 M KD value (AAV3) = <1.0E-12 M
Synomym	Adeno-associated virus 2; AAV-2
Conjugate	Unconjugated
Purification	Affinity chromatography
Storage before reconstitution	2-8°C until indicated expiry date
Storage after reconstitution	2-8°C
Intended use	Research use only
Application	Affinity chromatography, Dot blot, ELISA, ICC/IF, IP, Neutralization assay
Reactivity	AAV2, AAV2 7m8, AAV3
No reactivity	AAV1, AAV11, AAV12, AAV4, AAV5, AAV6, AAV7, AAV8, AAV9, AAVDJ, AAVrh10, AAVrh74

Applications

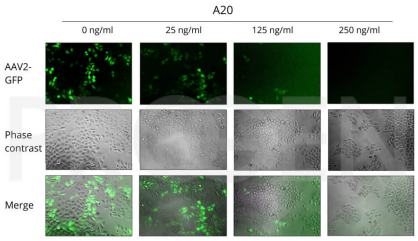
Affinity Chromatography	Assay dependent
Dot Blot	1:500 (0.1 µg/ml; non-denaturing conditions)
ELISA	Assay dependent
Immunocytochemistry (ICC)	1:20
Immunoprecipitation (IP)	1:5
Neutralization Assay	EC50 ~5 ng/ml (AAV2) and ~3 ng/ml (AAV3) - assay dependent

Background

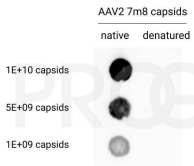
For characterization of different stages of infection and very useful for the analysis of the AAV2 assembly process. A20 specifically reacts with AAV2 and AAV3, empty and full capsids. Recognizes a conformational epitope of assembled capsids, not present in denatured capsid proteins and native but unassembled capsid proteins. The antibody cannot be used for immunoblotting.Epitope mapping experiments (Wobus et al. 2000) identified four immunoreactive (discontinous) regions. The major reaction was attributed to sequence aa 369 to aa378 of AAV2 capsids. The antibody is also useful for neutralizing experiments.

Wobus, C. E. et al. Monoclonal antibodies against the adeno-associated virus type 2 (AAV-2) capsid: epitope mapping and identification of capsid domains involved in AAV-2-cell interaction and neutralization of AAV-2 infection. J. Virol. 74, 9281-93 (2000). Moskalenko, M. et al. Epitope Mapping of Human Anti-Adeno-Associated Virus Type 2 Neutralizing Antibodies: Implications for Gene Therapy and Virus Structure. Jounal Virol. 74, 1761-1766 (2000).

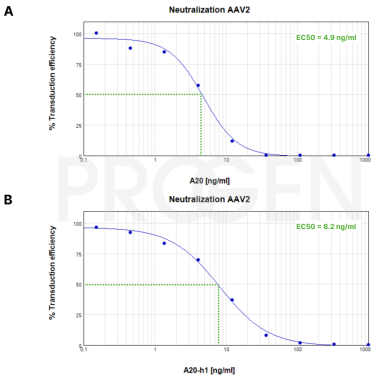
Product images



Neutralization of AAV2-GFP vectors with the A20 antibody (Cat. No. 61055). AAV infection was shown in HeLa cells and photos (GFP, CPE, merge) were taken ~48 h post infection. Neutralization was enhanced with increasing A20 concentration.



Dot blot analysis of native AAV2 7m8 capsids (1E+09-1E+10 capsids) and denatured AAV2 7m8 capsids (1E+09-1E+10 capsids, denatured for at 95°C for 10 min in sample buffer).The nitrocellulose membrane was blocked with 5% dry milk in PBST (PBS + 0.1% Tween 20) for 1 h at RT. The primary antibody anti-AAV2 (intact particle) mouse monoclonal, A20 (Cat. No. 61055) was diluted in blocking buffer (antibody concentration 100 ng/ml) and incubated for 1 h at RT. The secondary antibody goat anti-mouse IgG HRP was also diluted in blocking buffer (antibody concentration 200 ng/ml) and incubated for 1 h at RT. The bands were visualized by chemiluminescent detection using Pierce ECL Plus Western Blotting Substrate.



Neutralization of AAV2 with mouse monoclonal AAV2 antibody clone A20 (A) and human chimeric AAV2 antibody clone A20-h1 (B) by using AAV2-NanoLuc® viral particles from Promega. (A) anti-AAV2 (intact particle) mouse monoclonal, A20 (Cat. No. 61055) or (B) anti-AAV2, human chimeric, A20-h1 (Cat. No. 692379) were preincubated with AAV2-NanoLuc® viral particles for 30 min at RT at 300 rpm (antibody concentrations 0.2-3,000 ng/ml). HEK293 cells (100 µl) were plated at 200,000 cells/ml in DMEM + 1% FCS. Virus-antibody-mix (20 µl) was added to the cells and incubated for 16-24 h at 37°C. Extracellular NanoLuc Inhibitor and Nano-Glo® Live Cell Assay System (Promega) was added to the wells and incubated for 5 min at RT at 300 rpm. Luminescence was measured using an ID5-Reader and plotted with Softmax Pro 7.1 software to determine the EC50 values.

References

Publication	Species	Application
Wistuba, A., Weger, S., Kern, A., Rgen, J. & Kleinschmidt, A. Intermediates of Adeno-Associated Virus Type 2 Assembly: Identification of Soluble Complexes Containing Rep and Cap Proteins. J. Virol. 69, 5311â€“5319 (1995).	AAV2	IP
Moskalenko, M. et al. Epitope mapping of Human Anti-Adeno-Associated Virus Type 2 Neutralizing Antibodies: Implications for Gene Therapy and Virus Structure. Jounal Virol. 74, 1761â€“1766 (2000).	AAV2	neutralization, epitope mapping
McCraw, D. M., Oâ€™Donnell, J. K., Taylor, K. A., Stagg, S. M. & Chapman, M. S. Structure of adeno-associated virus-2 in complex with neutralizing monoclonal antibody A20. Virology 431, 40â€“49 (2012).	AAV2	cryoEM
McCraw, D. M., Oâ€™Donnell, J. K., Taylor, K. A., Stagg, S. M. & Chapman, M. S. Structure of adeno-associated virus-2 in complex with neutralizing monoclonal antibody A20. Virology 431, 40â€“49 (2012).	AAV2	cryoEM
Huttner, N. A. et al. Genetic modifications of the adeno-associated virus type 2 capsid reduce the affinity and the neutralizing effects of human serum antibodies. Gene Ther. 10, 2139â€“2147 (2003).	AAV2	epitope mapping